

The Proximal Origin of HCoV-19

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Since the first reports of novel pneumonia (COVID-19) in Wuhan, Hubei province, China^{1,2} there has been considerable discussion on the origin of the causative virus, human coronavirus 2019 (HCoV-19³; also referred to as SARS-CoV-2⁴). Infections with HCoV-19 are now widespread, and as of 29 February 2020, 86,012 cases have been confirmed in 57 countries, with 2,941 deaths⁵, although these are likely an underestimate with limited reporting of mild and asymptomatic cases.

HCoV-19 is the seventh coronavirus known to infect humans. Three of these viruses, SARS-CoV, MERS-CoV, and HCoV-19 can cause severe disease; four, HKU1, NL63, OC43 and 229E, are associated with mild respiratory symptoms⁷. Herein, we review what can be deduced about the origin of HCoV-19 from the comparative analysis of genome sequence data. In particular, we offer a perspective on the notable features in the HCoV-19 genome and discuss scenarios by which they could have arisen. Our analysis provides strong evidence that HCoV-19 is not a laboratory construct nor a purposefully manipulated virus.

Notable features of the HCoV-19 genome

Our genomic comparison of alpha- and betacoronaviruses (family *Coronaviridae*) identifies two notable features of the HCoV-19 genome: *(i)* based on structural studies⁸⁻¹⁰ and biochemical experiments^{1,9,11,12}, HCoV-19 appears optimized for binding to the human ACE2 receptor; *(ii)* the highly variable spike (S) protein of HCoV-19 has a functional polybasic (furin) cleavage site at the S1/S2 boundary through the insertion of twelve nucleotides^{10,13,14}. Additionally, this led to the predicted acquisition of three O-linked glycans around the polybasic cleavage site.

Mutations in the receptor binding domain of HCoV-19

The receptor binding domain (RBD) in the spike protein is the most variable part of the coronavirus genome^{1,2}. Six RBD amino acids have been shown to be critical for binding to ACE2 receptors and determining the host range of SARS-like viruses⁸. Using coordinates based SARS-CoV, they are Y442, L472, N479, D480, T487, and Y4911 corresponding to L455, F486, Q493, S494, N501, and Y505 in HCoV-19⁸. Five of these six residues differ between HCoV-19 and SARS-CoV (Fig. 1a). Based on structural studies⁸⁻¹⁰ and biochemical experiments^{1,9,11,12}, HCoV-19 seems to have an RBD that binds with high affinity to ACE2 from human, non-human primate, ferret, pig, and cat, and some other species with high receptor homology⁸.

While these analyses suggest that HCoV-19 may be capable of binding human ACE2 with high affinity, computational analyses predict that the interaction is not ideal⁸ and the RBD sequence is different from those shown in SARS-CoV to be optimal for receptor binding¹⁶. Thus, the optimized binding of HCoV-19 spike protein to human ACE2 is most likely the result of natural selection on a human or human-like ACE2 permitting another optimal binding solution to arise. This is strong evidence that HCoV-19 is *not* the product of purposeful manipulation.

Polybasic furin cleavage site and O-linked glycans

The second notable feature of HCoV-19 is a polybasic furin cleavage site (RRAR) at the junction of S1 and S2, the two subunits of the spike (**Fig. 1b**)^{13,14}. This allows effective cleavage by furin and other proteases and plays an important role in determining virus infectivity and host range¹⁸. In addition, a leading proline is also inserted at this site in HCoV-19; thus, the inserted sequence is PRRA (**Fig. 1b**). The strong turn created by the proline is predicted to result in the addition of O-linked glycans to S673, T678, and S686 flanking the cleavage site that are unique to HCoV-19 (**Fig. 1b**). Polybasic cleavage sites have not been observed in related “lineage B” betacoronaviruses, although other human betacoronaviruses, including HKU1 (lineage A), have them and predicted O-linked glycans near the S1/S2 cleavage site¹⁹. Given the level of genetic variation in the S protein it is likely that HCoV-19-like viruses with partial or full polybasic sites will be discovered in other species.

The functional consequence of the furin cleavage site in HCoV-19 is unknown and it will be important to determine what impact the feature has on transmissibility and pathogenesis in animal models²¹. Experiments with SARS-CoV have shown that insertion of a furin cleavage site at the S1/S2 junction enhances cell-cell fusion without affecting virus entry²². In addition, efficient cleavage of the MERS-CoV spike enables MERS-like coronaviruses from bats to infect human cells²³. In avian influenza viruses, rapid virus replication and transmission in highly dense chicken populations selects for the acquisition of polybasic cleavage sites in the haemagglutinin (HA) protein, ²⁴⁻²⁶. HA serves a similar function in cell-cell fusion and viral entry as the coronavirus spike protein. Acquisition polybasic cleavage sites in HA, by insertion or recombination, converts low pathogenicity avian influenza viruses into highly pathogenic forms²⁴⁻²⁶. The acquisition of polybasic cleavage sites by HA has also been observed after repeated passage in cell culture or through animals^{27,28}.

The function of the predicted O-linked glycans is less clear, but they could create a “mucin-like domain” shielding potential epitopes or key residues on the HCoV-19 spike protein^{30,31}. Several viruses employ mucin-like domains as part of a glycan shield involved in immune evasion³⁰. Although prediction of O-linked glycosylation is robust³², biochemical analyses or structural studies are required to determine whether or not these sites in HCoV-19 are utilized.

Theories of HCoV-19 origins

It is improbable that HCoV-19 emerged through laboratory manipulation or engineering of a related SARS-like coronavirus. As noted above, the RBD of HCoV-19 is optimized for human ACE2 binding with an efficient solution different from those previously predicted^{8,16}. Further, had genetic manipulation had been performed, one of the several reverse genetic systems available for betacoronaviruses would likely have been used²⁰. However, the genetic data irrefutably show that HCoV-19 is not derived from any previously used virus backbone³³. Instead, we propose two scenarios that can plausibly explain the origin of HCoV-19: (i) natural selection in a non-human animal host prior to zoonotic transfer, and (ii) natural selection in humans following zoonotic transfer. We also discuss whether selection during passage in culture could have given rise to the same observed features.

Natural selection in an animal host prior to zoonotic transfer

As many of the early cases of COVID-19 were linked to the Huanan seafood and wildlife market in Wuhan^{1,2}, it is possible that an animal source was present at this location. Given the similarity of HCoV-19 to bat SARS-like coronaviruses², it is likely that bats serve as reservoir hosts for its progenitor. Although RaTG13, sampled from a *Rhinolophus affinis* bat, is ~96% identical overall to HCoV-19¹, its spike diverges in the RBD suggesting that it may not bind efficiently to the human ACE2 receptor (**Fig. 1a**)⁸.

Malayan pangolins (*Manis javanica*) illegally imported into Guangdong province contain coronaviruses similar to HCoV-19^{15,37-39}. Although the RaTG13 bat virus remains the closest relative to HCoV-19 across the whole genome¹, some pangolin coronaviruses exhibit strong similarity to HCoV-19 in the RBD, including all six key RBD residues (**Fig. 1**)^{15,39}. This clearly shows the HCoV-19 spike protein optimized for binding to human-like ACE2 occurs in nature and is the result of natural selection. Similarly, neither the bat nor pangolin betacoronaviruses sampled to date carry polybasic cleavage sites. Although a non-human animal coronavirus, sufficiently similar to HCoV-19 across its entire genome that it could have served as the direct progenitor of the virus, has yet to be identified, the diversity of coronaviruses in bats and other species is massively undersampled. Mutations, including point mutations, insertions and deletions, can occur near the S1/S2 junction of coronaviruses^{34,40-43} suggesting that the polybasic site could arise by a natural evolutionary process. For a precursor virus to acquire both the polybasic cleavage site and mutations in the spike protein suitable for human ACE2 receptor binding, an animal host would likely have to have a high population density – to allow natural selection to proceed efficiently – and an ACE2 gene that is similar to the human orthologue.

Natural selection in humans following zoonotic transfer

It is possible that a progenitor to HCoV-19 jumped into humans, acquiring the genomic features described above through adaptation during (undetected) human-to-human transmission. Once acquired, these adaptations would enable the epidemic to take off, producing a sufficiently large and unusual cluster of pneumonia cases to trigger the surveillance system that ultimately detected it^{1,2}.

All HCoV-19 genomes sequenced so far have the genomic features derived above and are thus derived from a common ancestor that had them too. The presence in pangolins of an RBD very similar to that in HCoV-19 means we can infer this was also likely in the virus that jumped to humans. This leaves the polybasic cleavage site insertion to occur during human-to-human transmission.

Estimates of the timing of the most recent common ancestor of HCoV-19 using currently available genome sequence data point to virus emergence in late November to early December 2019⁴⁴⁻⁴⁶, compatible with the earliest retrospectively confirmed cases⁴⁷. Hence, this scenario presumes a period of unrecognised transmission in humans between the initial zoonotic transfer event and the acquisition of the polybasic cleavage site. Sufficient opportunity could occur if there had been many prior zoonotic events producing short chains of human-to-human transmission over an extended period. This is essentially the situation for MERS-CoV in the Arabian Peninsula where all the human cases are the result of repeated jumps of the virus from dromedary camels, producing single infections or short transmission chains that eventually resolve, with no adaptation to sustained human transmission⁴⁸.

Metagenomic studies of banked human samples could provide important information on whether this cryptic spread has occurred, although given the relatively short period of viremia it may be impossible to detect low level HCoV-19 circulation in historical samples. Retrospective serological studies could also be informative and a few such studies have been conducted. One found that animal importation traders had a 13% seropositivity to coronaviruses⁴⁹, while another noted that 3% residents of a village in Southern China were seropositive to SARS-like coronaviruses⁵⁰. Critically, however, these studies could not have distinguished whether positive serological responses were due to prior infections with SARS-CoV, HCoV-19,

or other SARS-like coronaviruses. Further serological studies should be conducted to determine the extent of prior human exposure to HCoV-19.

Selection during passage

Basic research involving passage of bat SARS-like coronaviruses in cell culture and/or animal models have been ongoing in BSL-2 for many years in laboratories across the world⁵¹⁻⁵⁴. There are also several documented instances of laboratory escapes of SARS-CoV⁵⁵⁻⁵⁷. We must therefore examine the possibility of an inadvertent laboratory release of HCoV-19.

In theory, it is possible that HCoV-19 acquired RBD mutations (**Fig. 1a**) during adaptation to passage in cell culture, as has been observed in studies with SARS-CoV¹⁶ and MERS-CoV⁵⁸. The finding of SARS-like coronaviruses from pangolins with near-identical RBDs, however, provides a much stronger and parsimonious explanation for how HCoV-19 acquired these via recombination or mutation²⁰.

The acquisition of both the polybasic cleavage site and predicted O-linked glycans also argues against any type of culture-based scenario. New polybasic cleavage sites have only been observed after prolonged passage of low pathogenicity avian influenza virus *in vitro* or *in vivo*^{24,26-28}. Furthermore, a hypothetical generation of HCoV-19 by cell culture or animal passage would have required prior isolation of a progenitor virus with a very high genetic similarity, which has not been described. Subsequent generation of a polybasic cleavage site would have then required repeated passage in cell culture or animals with ACE2 receptors similar to humans (e.g. ferrets), but such work has also not previously been described. Finally, the generation of the predicted O-linked glycans is also unlikely to have occurred due to cell culture passage, as such features suggest the involvement of an immune system³⁰.

Conclusions

In the midst of the global COVID-19 public health emergency it is reasonable to wonder why the origins of the epidemic matter. A detailed understanding of how an animal virus jumped species boundaries to infect humans so productively will help in the prevention of future zoonotic events. For example, if HCoV-19 pre-adapted in another animal species then we are at risk of future re-emergence events. In contrast, if the adaptive process we describe occurred in humans, then even if we have repeated zoonotic transfers they are unlikely to take-off without the same series of mutations. In addition, identifying the closest animal relatives of HCoV-19 will greatly assist studies of virus function. Indeed, the availability of the RaTG13 bat sequence helped reveal the key mutations in the RBD as well as the polybasic cleavage site insertion.

The genomic features described here may in part explain the infectiousness and transmissibility of HCoV-19 in humans. Although the evidence shows that HCoV-19 is not a purposefully manipulated virus, it is currently impossible to prove or disprove the other theories of its origin described here. However, since we observe all notable HCoV-19 features - including the optimized RBD and furin cleavage site - in related coronaviruses in nature, we do not believe that selection during passage, or any other type of laboratory-based scenario, is necessary.

More scientific data could swing the balance of evidence to favor one hypothesis over another. Obtaining virus sequences from any immediate non-human animal source would be the most definitive way of revealing virus origins. For example, a future observation of an intermediate or fully formed polybasic cleavage site in an HCoV-19 related virus from animals would lend very strong support to the natural selection hypotheses. It would also be helpful to obtain more genetic and functional data about HCoV-19, including experimental studies of receptor binding and the role of the polybasic cleavage site and predicted O-linked glycans. The identification of a potential intermediate host of HCoV-19, as well as the sequencing of very early cases would similarly be highly informative. Irrespective of the exact mechanisms of how

HCoV-19 originated via natural selection, the ongoing surveillance of pneumonia in humans and other animals is clearly of utmost importance.

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Figure Legends

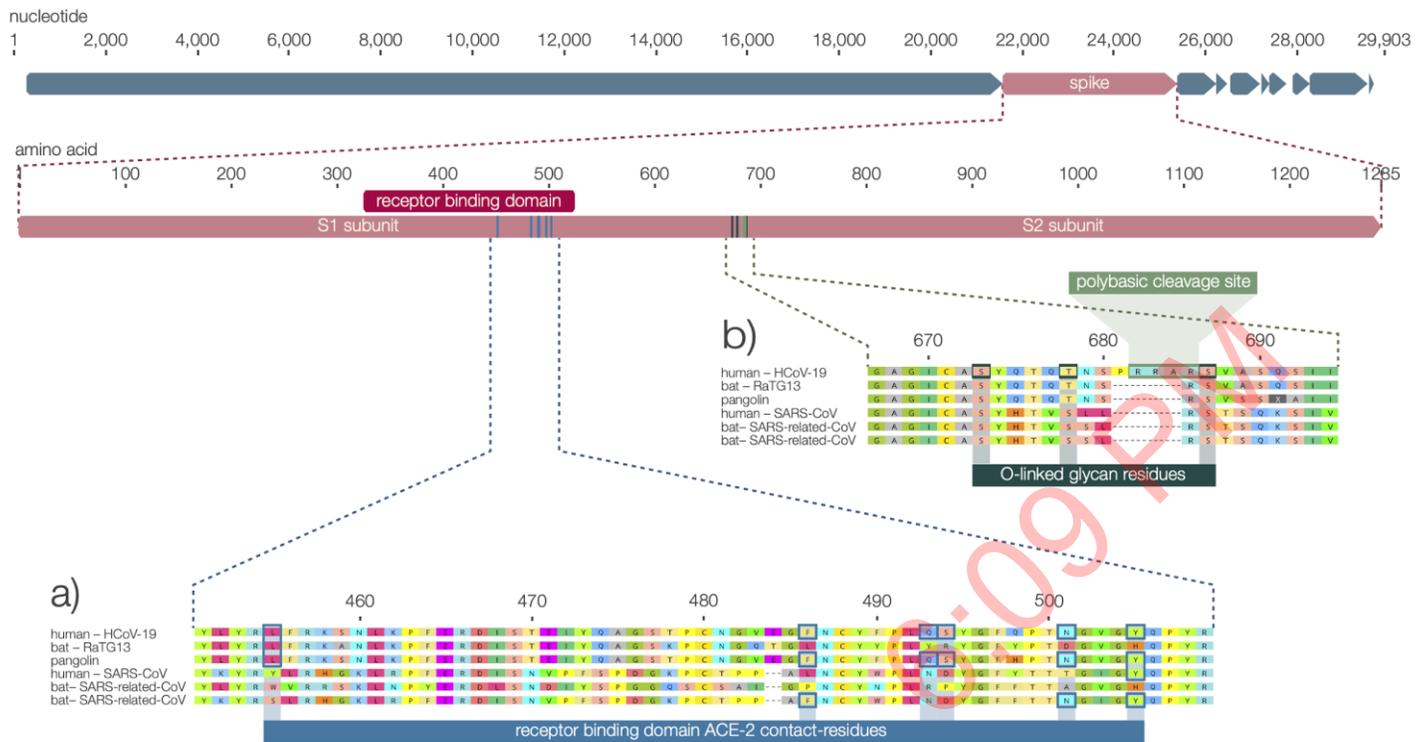


Figure 1. (a) Mutations in contact residues of the HCoV-19 spike protein. The spike protein of HCoV-19 (top) was aligned against the most closely related SARS-like CoVs and SARS-CoV. Key residues in the spike protein that make contact to the ACE2 receptor are marked with blue boxes in both HCoV-19 and the SARS-CoV Urbani strain. (b) Acquisition of polybasic cleavage site and O-linked glycans. Both the polybasic cleavage site and the three adjacent predicted O-linked glycans are unique to HCoV-19 and not previously seen in lineage B betacoronaviruses. Sequences shown are from NCBI GenBank, accession numbers MN908947, MN996532, AY278741, KY417146 and MK211376. The pangolin coronavirus sequences are a consensus generated from SRR10168377 and SRR10168378 (NCBI BioProject PRJNA573298)^{37,59}.

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References

1. Zhou, P. *et al.* A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* (2020) doi:10.1038/s41586-020-2012-7.
2. Wu, F. *et al.* A new coronavirus associated with human respiratory disease in China. *Nature* (2020) doi:10.1038/s41586-020-2008-3.
3. Jiang, S. *et al.* A distinct name is needed for the new coronavirus. *Lancet* (2020) doi:10.1016/S0140-6736(20)30419-0.
4. Gorbalenya, A. E. Severe acute respiratory syndrome-related coronavirus—The species and its viruses, a statement of the Coronavirus Study Group. *BioRxiv* (2020).
5. Dong, E., Du, H. & Gardner, L. An interactive web-based dashboard to track COVID-19 in real time. *Lancet Infect. Dis.* (2020) doi:10.1016/S1473-3099(20)30120-1.
6. Statement on the second meeting of the International Health Regulations (2005) Emergency Committee regarding the outbreak of novel coronavirus (2019-nCoV). [https://www.who.int/news-room/detail/30-01-2020-statement-on-the-second-meeting-of-the-international-health-regulations-\(2005\)-emergency-committee-regarding-the-outbreak-of-novel-coronavirus-\(2019-ncov\)](https://www.who.int/news-room/detail/30-01-2020-statement-on-the-second-meeting-of-the-international-health-regulations-(2005)-emergency-committee-regarding-the-outbreak-of-novel-coronavirus-(2019-ncov)).
7. Corman, V. M., Muth, D., Niemeyer, D. & Drosten, C. Hosts and Sources of Endemic Human Coronaviruses. *Adv. Virus Res.* **100**, 163–188 (2018).
8. Wan, Y., Shang, J., Graham, R., Baric, R. S. & Li, F. Receptor recognition by novel coronavirus from Wuhan: An analysis based on decade-long structural studies of SARS. *J. Virol.* (2020) doi:10.1128/JVI.00127-20.
9. Wrapp, D. *et al.* Cryo-EM Structure of the 2019-nCoV Spike in the Prefusion Conformation. *bioRxiv* 2020.02.11.944462 (2020) doi:10.1101/2020.02.11.944462.
10. Walls, A. C. *et al.* Structure, function and antigenicity of the SARS-CoV-2 spike glycoprotein. *bioRxiv* 2020.02.19.956581 (2020) doi:10.1101/2020.02.19.956581.

11. Letko, M., Marzi, A. & Munster, V. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. *Nat Microbiol* (2020) doi:10.1038/s41564-020-0688-y.
12. Hoffmann, M. *et al.* The novel coronavirus 2019 (2019-nCoV) uses the SARS-coronavirus receptor ACE2 and the cellular protease TMPRSS2 for entry into target cells. *bioRxiv* 2020.01.31.929042 (2020) doi:10.1101/2020.01.31.929042.
13. Gallaher, W. Analysis of Wuhan Coronavirus: Deja Vu. *Virological* <http://virological.org/t/analysis-of-wuhan-coronavirus-deja-vu/357> (2020).
14. Coutard, B. *et al.* The spike glycoprotein of the new coronavirus 2019-nCoV contains a furin-like cleavage site absent in CoV of the same clade. *Antiviral Res.* 104742 (2020).
15. Zhang, T., Wu, Q. & Zhang, Z. Pangolin homology associated with 2019-nCoV. *bioRxiv* 2020.02.19.950253 (2020) doi:10.1101/2020.02.19.950253.
16. Sheahan, T. *et al.* Mechanisms of zoonotic severe acute respiratory syndrome coronavirus host range expansion in human airway epithelium. *J. Virol.* **82**, 2274–2285 (2008).
17. Calisher, C. *et al.* Statement in support of the scientists, public health professionals, and medical professionals of China combatting COVID-19. *Lancet* (2020) doi:10.1016/S0140-6736(20)30418-9.
18. Nao, N. *et al.* Genetic Predisposition To Acquire a Polybasic Cleavage Site for Highly Pathogenic Avian Influenza Virus Hemagglutinin. *MBio* **8**, (2017).
19. Chan, C.-M. *et al.* Spike protein, S, of human coronavirus HKU1: role in viral life cycle and application in antibody detection. *Exp. Biol. Med.* **233**, 1527–1536 (2008).
20. Cui, J., Li, F. & Shi, Z.-L. Origin and evolution of pathogenic coronaviruses. *Nat. Rev. Microbiol.* **17**, 181–192 (2019).
21. Bao, L. *et al.* The Pathogenicity of SARS-CoV-2 in hACE2 Transgenic Mice. *bioRxiv* 2020.02.07.939389 (2020) doi:10.1101/2020.02.07.939389.
22. Follis, K. E., York, J. & Nunberg, J. H. Furin cleavage of the SARS coronavirus spike glycoprotein

- enhances cell-cell fusion but does not affect virion entry. *Virology* **350**, 358–369 (2006).
23. Menachery, V. D. *et al.* Trypsin treatment unlocks barrier for zoonotic bat coronaviruses infection. *J. Virol.* (2019) doi:10.1128/JVI.01774-19.
 24. Longping, V. T., Hamilton, A. M., Friling, T. & Whittaker, G. R. A novel activation mechanism of avian influenza virus H9N2 by furin. *J. Virol.* **88**, 1673–1683 (2014).
 25. Alexander, D. J. & Brown, I. H. History of highly pathogenic avian influenza. *Rev. Sci. Tech.* **28**, 19–38 (2009).
 26. Luczo, J. M. *et al.* Evolution of high pathogenicity of H5 avian influenza virus: haemagglutinin cleavage site selection of reverse-genetics mutants during passage in chickens. *Sci. Rep.* **8**, 11518 (2018).
 27. Ito, T. *et al.* Generation of a highly pathogenic avian influenza A virus from an avirulent field isolate by passaging in chickens. *J. Virol.* **75**, 4439–4443 (2001).
 28. Li, S. Q., Orlich, M. & Rott, R. Generation of seal influenza virus variants pathogenic for chickens, because of hemagglutinin cleavage site changes. *J. Virol.* **64**, 3297–3303 (1990).
 29. Shengqing, Y. *et al.* Generation of velogenic Newcastle disease viruses from a nonpathogenic waterfowl isolate by passaging in chickens. *Virology* **301**, 206–211 (2002).
 30. Bagdonaite, I. & Wandall, H. H. Global aspects of viral glycosylation. *Glycobiology* **28**, 443–467 (2018).
 31. Tran, E. E. H. *et al.* Spatial localization of the Ebola virus glycoprotein mucin-like domain determined by cryo-electron tomography. *J. Virol.* **88**, 10958–10962 (2014).
 32. Steentoft, C. *et al.* Precision mapping of the human O-GalNAc glycoproteome through SimpleCell technology. *EMBO J.* **32**, 1478–1488 (2013).
 33. Almazán, F. *et al.* Coronavirus reverse genetic systems: infectious clones and replicons. *Virus Res.* **189**, 262–270 (2014).
 34. Guan, Y. *et al.* Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. *Science* **302**, 276–278 (2003).

35. Haagmans, B. L. *et al.* Middle East respiratory syndrome coronavirus in dromedary camels: an outbreak investigation. *Lancet Infect. Dis.* **14**, 140–145 (2014).
36. Azhar, E. I. *et al.* Evidence for camel-to-human transmission of MERS coronavirus. *N. Engl. J. Med.* **370**, 2499–2505 (2014).
37. Wong, M. C., Javornik Cregeen, S. J., Ajami, N. J. & Petrosino, J. F. Evidence of recombination in coronaviruses implicating pangolin origins of nCoV-2019. *bioRxiv* 2020.02.07.939207 (2020) doi:10.1101/2020.02.07.939207.
38. Lam, T. T.-Y. *et al.* Identification of 2019-nCoV related coronaviruses in Malayan pangolins in southern China. *bioRxiv* (2020) doi:10.1101/2020.02.13.945485.
39. Xiao, K. *et al.* Isolation and Characterization of 2019-nCoV-like Coronavirus from Malayan Pangolins. *bioRxiv* 2020.02.17.951335 (2020) doi:10.1101/2020.02.17.951335.
40. Licitra, B. N. *et al.* Mutation in spike protein cleavage site and pathogenesis of feline coronavirus. *Emerg. Infect. Dis.* **19**, 1066–1073 (2013).
41. Yamada, Y. & Liu, D. X. Proteolytic activation of the spike protein at a novel RRRR/S motif is implicated in furin-dependent entry, syncytium formation, and infectivity of coronavirus infectious bronchitis virus in cultured cells. *J. Virol.* **83**, 8744–8758 (2009).
42. Yamada, Y. K., Takimoto, K., Yabe, M. & Taguchi, F. Requirement of proteolytic cleavage of the murine coronavirus MHV-2 spike protein for fusion activity. *Adv. Exp. Med. Biol.* **440**, 89–93 (1998).
43. Lamers, M. M. *et al.* Deletion Variants of Middle East Respiratory Syndrome Coronavirus from Humans, Jordan, 2015. *Emerg. Infect. Dis.* **22**, 716–719 (2016).
44. Phylodynamic Analysis | 90 genomes | 12 Feb 2020 – Novel 2019 coronavirus / nCoV-2019 Genomic Epidemiology. <http://virological.org/t/phylodynamic-analysis-90-genomes-12-feb-2020/356> (2020).
45. Phylodynamic estimation of incidence and prevalence of novel coronavirus (nCoV) infections through time – Novel 2019 coronavirus / nCoV-2019 Genomic Epidemiology.

- <http://virological.org/t/phylogenetic-estimation-of-incidence-and-prevalence-of-novel-coronavirus-nCoV-infections-through-time/391> (2020).
46. Clock and TMRCA based on 27 genomes. *Virological* <http://virological.org/t/clock-and-tmrca-based-on-27-genomes/347> (2020).
 47. Huang, C. *et al.* Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* (2020) doi:10.1016/S0140-6736(20)30183-5.
 48. Dudas, G., Carvalho, L. M., Rambaut, A. & Bedford, T. MERS-CoV spillover at the camel-human interface. *Elife* **7**, (2018).
 49. Centers for Disease Control and Prevention (CDC). Prevalence of IgG antibody to SARS-associated coronavirus in animal traders--Guangdong Province, China, 2003. *MMWR Morb. Mortal. Wkly. Rep.* **52**, 986–987 (2003).
 50. Wang, N. *et al.* Serological Evidence of Bat SARS-Related Coronavirus Infection in Humans, China. *Virol. Sin.* **33**, 104–107 (2018).
 51. Ge, X.-Y. *et al.* Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor. *Nature* **503**, 535–538 (2013).
 52. Hu, B. *et al.* Discovery of a rich gene pool of bat SARS-related coronaviruses provides new insights into the origin of SARS coronavirus. *PLoS Pathog.* **13**, e1006698 (2017).
 53. Zeng, L.-P. *et al.* Bat Severe Acute Respiratory Syndrome-Like Coronavirus WIV1 Encodes an Extra Accessory Protein, ORFX, Involved in Modulation of the Host Immune Response. *J. Virol.* **90**, 6573–6582 (2016).
 54. Yang, X.-L. *et al.* Isolation and Characterization of a Novel Bat Coronavirus Closely Related to the Direct Progenitor of Severe Acute Respiratory Syndrome Coronavirus. *J. Virol.* **90**, 3253–3256 (2015).
 55. Lim, P. L. *et al.* Laboratory-acquired severe acute respiratory syndrome. *N. Engl. J. Med.* **350**, 1740–1745 (2004).

56. Senior, K. Recent Singapore SARS case a laboratory accident. *Lancet Infect. Dis.* **3**, 679 (2003).
57. Lim, W., Ng, K.-C. & Tsang, D. N. C. Laboratory containment of SARS virus. *Ann. Acad. Med. Singapore* **35**, 354–360 (2006).
58. Letko, M. *et al.* Adaptive Evolution of MERS-CoV to Species Variation in DPP4. *Cell Rep.* **24**, 1730–1737 (2018).
59. Liu, P., Chen, W. & Chen, J.-P. Viral Metagenomics Revealed Sendai Virus and Coronavirus Infection of Malayan Pangolins (*Manis javanica*). *Viruses* vol. 11 979 (2019).

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