

RCC - CCR STUDY NUMBER 1061403

**SALMONELLA TYPHIMURIUM
AND
ESCHERICHIA COLI
REVERSE MUTATION ASSAY**

WITH

Glyphosate technical (NUP-05067)

FINAL REPORT

**STUDY COMPLETION DATE:
MARCH 16, 2007**



1 STATEMENT OF NO DATA CONFIDENTIALITY CLAIM

No claim of confidentiality is made for any information contained in this study on the basis of it falling within the scope of FIFRA Sect. 10 (d)(1)(A), (B) or (C).

Company

Company Agent

Title

Signature

Date

These data are considered to be CONFIDENTIAL for all purposes other than compliance with FIFRA Section 10. Submission of these data in compliance with FIFRA does not constitute a waiver of any right to confidentiality, which may exist under any statute or in any other country.

The above statement supersedes any other markings of confidentiality which may appear elsewhere in the report.

2 CONTENTS

1	STATEMENT OF NO DATA CONFIDENTIALITY CLAIM	2
2	CONTENTS	3
3	PREFACE	4
3.1	General	4
3.2	Responsibilities	4
3.3	Schedule	4
3.4	Project Staff Signatures	5
3.5	Good Laboratory Practice	5
3.6	Guidelines	5
3.7	Archiving	6
3.8	Deviations to Study Plan	6
4	STATEMENT OF COMPLIANCE	7
5	STATEMENT OF QUALITY ASSURANCE UNIT	8
6	SUMMARY OF RESULTS	9
6.1	Conclusion	9
7	OBJECTIVE	10
7.1	Aims of the Study	10
7.2	Reasons for the Study	10
8	MATERIALS AND METHODS	11
8.1	Test Item	11
8.2	Controls	12
8.3	Test System	13
8.4	Mammalian Microsomal Fraction S9 Mix	15
8.5	Pre-Experiment for Toxicity	16
8.6	Dose Selection	16
8.7	Experimental Performance	16
8.8	Data Recording	17
8.9	Acceptability of the Assay	17
8.10	Evaluation of Results	17
8.11	Biometry	17
9	DISCUSSION OF RESULTS	18
10	REFERENCES	19
11	DISTRIBUTION OF THE REPORT	19
12	SUMMARY OF RESULTS	20
12.1	Summary of Results Pre-Experiment/Experiment I	20
12.2	Summary of Results Experiment II	21
13	HISTORICAL CONTROL DATA	22
14	ANNEX I: TABLES OF RESULTS (8 PAGES)	23
15	ANNEX II: COPY OF GLP CERTIFICATE OF ANALYSIS	32
16	ANNEX III: COPY OF GLP CERTIFICATE	33

3 PREFACE

3.1 General

Title: Salmonella typhimurium and Escherichia coli
Reverse Mutation Assay with Glyphosate technical
(NUP-05067)

Sponsor: Nufarm Asia Sdn Bhd
No. 9A & B, Jalan USJ 21/5
47630 Subang Jaya, Selangor, D.E.
Malaysia

Study Monitor: Dr. [REDACTED]
Nufarm Asia Sdn Bhd

Test Facility: R C C
Cytotest Cell Research GmbH (RCC-CCR)
In den Leppsteinswiesen 19
64380 Rossdorf
Germany

Contracting Institute: R C C Ltd
4452 Ittingen
Switzerland

Reference Number: B02518

3.2 Responsibilities

Study Director: Dipl. Biol. [REDACTED]

Deputy Study Director: Dr. [REDACTED]

Management: Dr. [REDACTED]

Head of Quality Assurance Unit: [REDACTED]

3.3 Schedule

Date of the Study Plan: January 03, 2007

Experimental Starting Date: January 16, 2007

Experimental Completion Date: January 25, 2007

Date of Draft Report: February 23, 2007

Date of Final Report: March 16, 2007

3.7 Archiving

RCC Cytotest Cell Research GmbH will archive the following data for 15 years:

Raw data, study plan, final report, and a sample of the test item.

No data will be discarded without the sponsor's consent.

3.8 Deviations to Study Plan

There were no deviations to study plan.

This document is not the property of EFSA and is provided for giving full effect to the right of public access to documents under EU law.
The document may be subject to rights such as intellectual property and copy rights of third parties.
Furthermore, this document may fall under a regulatory data protection regime.
Consequently, any publication, distribution, reproduction and/or publishing and any commercial exploitation and use
of this document may therefore be prohibited and violate the rights of its owner.

3.4 Project Staff Signatures

Study Director

Dipl. Biol. [redacted]

[redacted]
Date: March 16, 2007

Management

[redacted]
Date: March 16, 2007

3.5 Good Laboratory Practice

The study was performed in compliance with:

"Chemikaliengesetz" (Chemicals Act) of the Federal Republic of Germany, "Anhang 1" (Annex 1) dated July 25, 1994 ("BGBl. I 1994", pp. 1703), last revision dated June 27, 2002.

"OECD Principles of Good Laboratory Practice", as revised in 1997 [C(97)186/Final].

These procedures are consistent with Good Laboratory Practice Regulations specified by regulatory authorities throughout the European Community, the United States (EPA and FDA), and Japan (MHW, MAFF, and MITI).

3.6 Guidelines

This study followed the procedures indicated by the following internationally accepted guidelines and recommendations:

The OECD Guidelines for Testing of Chemicals No. 471: "Bacterial Reverse Mutation Test", adopted July 21, 1997 referenced as Method B13/14 of Commission Directive 92/69/EEC (which constitutes Annex V of Council Directive 67/548/EEC).

Japanese Ministry of Agriculture, Forestry and Fisheries (JMAFF), Guidelines for Study Results, Reverse mutation studies. Guideline No.2-1-19-1. >Notification 12NohSan No. 8147, as partly revised in 16-Shouan-9260, on March 16 2005. English translation by ACIS on October 17, 2005.

4 STATEMENT OF COMPLIANCE

Study Number: 1061403
Test Item: Glyphosate technical (NUP-05067)
Study Director: Dipl. Biol. [REDACTED]
Title: Salmonella typhimurium and Escherichia coli
Reverse Mutation Assay with Glyphosate technical
(NUP-05067)

This study performed in the test facility of RCC Cytotest Cell Research GmbH was conducted in compliance with Good Laboratory Practice Regulations:

"Chemikaliengesetz" (Chemicals Act) of the Federal Republic of Germany, "Anhang 1" (Annex 1) dated July 25, 1994 ("BGBl. I 1994", pp. 1703), last revision dated June 27, 2002.

"OECD Principles of Good Laboratory Practice", as revised in 1997 [C(97)186/Final]

There were no circumstances that may have affected the quality or integrity of the study.

Study Director

RCC - CCR

Dipl. Biol. [REDACTED]

[REDACTED]
Date: March 16, 2007

5 STATEMENT OF QUALITY ASSURANCE UNIT

Study Number: 1061403

Test Item: Glyphosate technical (NUP-05067)

Study Director: Dipl. Biol. [REDACTED]

Title: Salmonella typhimurium and Escherichia coli
Reverse Mutation Assay with Glyphosate technical
(NUP-05067)

The general facilities and activities of RCC Cytotest Cell Research GmbH are inspected periodically and the results are reported to the responsible person and the management.

Study procedures were inspected periodically. The study plan and this report were audited by the Quality Assurance Unit. The dates are given below.

Phases and Dates of QAU Inspections/ Audits		Dates of Reports to the Study Director and to Management
Study Plan (Draft):	January 03, 2007	January 03, 2007
Study Plan:	January 04, 2007	- -
Study Inspection:	January 16, 2007	January 16, 2007
Draft Report:	March 09, 2007	March 09, 2007

This statement is to confirm that the present final report reflects the raw data.

Head of Quality Assurance Unit

[REDACTED]

[REDACTED]

Date: March 16, 2007

6 SUMMARY OF RESULTS

This study was performed to investigate the potential of Glyphosate technical (NUP-05067) to induce gene mutations in the plate incorporation test (experiment I) and the pre-incubation test (experiment II) using the *Salmonella typhimurium* strains TA 1535, TA 1537, TA 98, and TA 100, and the *Escherichia coli* strain WP2 uvrA.

The assay was performed in two independent experiments both with and without liver microsomal activation. Each concentration, including the controls, was tested in triplicate. The test item was tested at the following concentrations:

Pre-Experiment/Experiment I: 3; 10; 33; 100; 333; 1000; 2500; and 5000 µg/plate

Experiment II: 33; 100; 333; 1000; 2500; and 5000 µg/plate

The plates incubated with the test item showed normal background growth in experiment I and II with and without metabolic activation. Only in experiment I in strain WP2 uvrA reduced background growth was observed at 5000 µg/plate in the absence of metabolic activation.

No toxic effects, evident as a reduction in the number of revertants, occurred in the test groups with and without metabolic activation. Minor toxic effects occurred at 5000 µg/plate in strain WP2 uvrA in the absence of metabolic activation in experiment I and II.

No substantial increase in revertant colony numbers of any of the five tester strains was observed following treatment with Glyphosate technical (NUP-05067) at any dose level, neither in the presence nor absence of metabolic activation (S9 mix). There was also no tendency of higher mutation rates with increasing concentrations in the range below the generally acknowledged border of biological relevance.

Appropriate reference mutagens were used as positive controls and showed a distinct increase of induced revertant colonies.

6.1 Conclusion

In conclusion, it can be stated that during the described mutagenicity test and under the experimental conditions reported, the test item did not induce gene mutations by base pair changes or frameshifts in the genome of the strains used.

Therefore, Glyphosate technical (NUP-05067) is considered to be non-mutagenic in this *Salmonella typhimurium* and *Escherichia coli* reverse mutation assay.

7 OBJECTIVE

7.1 Aims of the Study

The experiments were performed to assess the potential of the test item to induce gene mutations by means of two independent *Salmonella typhimurium* and *Escherichia coli* reverse mutation assays. Experiment I was performed as a plate incorporation assay. Since a negative result was obtained in this experiment, experiment II was performed as a pre-incubation assay.

7.2 Reasons for the Study

The most widely used assays for detecting gene mutations are those using bacteria (3). They are relatively simple and rapid to perform, and give reliable data on the ability of an agent to interact with DNA and produce mutations.

Reverse mutation assays determine the frequency with which an agent reverses or suppresses the effect of the forward mutation. The genetic target presented to an agent is therefore small, specific and selective. Several bacterial strains, or a single strain with multiple markers are necessary to overcome the effects of mutagen specificity. The reversion of bacteria from growth-dependence on a particular amino acid to growth in the absence of that amino acid (reversion from auxotrophy to prototrophy) is the most widely used marker.

The *Salmonella typhimurium* histidine (*his*) and the *E. coli* tryptophan (*trp*) reversion system measures $his^- \rightarrow his^+$ and $trp^- \rightarrow trp^+$ reversions, respectively. The *S. typhimurium* and *Escherichia coli* strains are constructed to differentiate between base pair (TA 1535, TA 100, and WP2 *uvrA*) and frameshift (TA 1537, TA 98) mutations.

According to the direct plate incorporation or the pre-incubation method the bacteria are exposed to the test item with and without metabolic activation and plated on selective medium. After a suitable period of incubation, revertant colonies are counted.

To establish a dose response effect at least 6 dose levels with adequately spaced intervals were tested. The maximum dose level was 5000 µg/plate.

To validate the test, reference mutagens were tested in parallel to the test item.

8 MATERIALS AND METHODS

8.1 Test Item

Internal RCC-CCR Test Item Number: S 693811

The test item and the information concerning the test item were provided by the sponsor.

Identity: Glyphosate technical (NUP-05067)

Batch No.: 0609-1

Aggregate state at room temperature: Crystalline powder

Colour: White

Purity: 95.0 %

Solubility: 10.9 g/L in water

Stability in water (sterile): 30 days at room temperature

Storage: Room temperature

Expiration Date: August 15, 2008

On the day of the experiment, the test item Glyphosate technical (NUP-05067) was dissolved in deionised water, the stock solution was neutralized with 5N sodium hydroxide. The solvent was chosen because of its solubility properties (5).

No precipitation of the test item occurred up to the highest investigated dose.

8.2 Controls

8.2.1 The Negative Controls

Concurrent untreated and solvent controls will be performed.

8.2.2 The Positive Control Substances

Without metabolic activation

Strains: TA 1535, TA 100
Name: Sodium azide, NaN_3
Supplier: SERVA, D-69042 Heidelberg
Lot Number: 14760
Catalogue No.: 30175
Purity: at least 99 %
Expiration Date: August 2007
Dissolved in: aqua deionised
Concentration: 10 µg/plate

Strains: TA 1537, TA 98
Name: 4-nitro-o-phenylene-diamine, 4-NOPD
Supplier: SIGMA, D-82041 Deisenhofen
Lot Number: 416324/1
Catalogue No.: N 9504
Purity: > 99.9 %
Expiration Date: April 2009
Dissolved in: DMSO (MERCK, D-64293 Darmstadt, purity > 99 %)
Concentration: 10 µg/plate in TA 98, 50 µg/plate in TA 1537

Strain: WP2 uvrA
Name: methyl methane sulfonate, MMS
Supplier: Merck-Schuchardt, D-85662 Hohenbrunn
Lot Number: 074K3720
Catalogue No.: 820775
Purity: > 99.0 %
Expiration Date: October 2007
Dissolved in: aqua deionised
Concentration: 3 µL/plate

With metabolic activation

Strains: TA 1535, TA 1537, TA 98, TA 100, WP2 uvrA
Name: 2-aminoanthracene, 2-AA
Supplier: SIGMA, D-82041 Deisenhofen
Lot Number: S11804-252
Catalogue No.: A 1381
Purity: 97.5 %
Expiration Date: November 2007
Dissolved in: DMSO (MERCK, D-64293 Darmstadt, purity > 99 %)
Concentration: 2.5 µg/plate (TA 1535, TA 1537, TA 98, TA 100),
10 µg/plate (WP2 uvrA)

The stability of the positive control substances in solution is unknown but a mutagenic response in the expected range will be sufficient evidence of biological stability.

8.3 Test System

8.3.1 Characterisation of the *Salmonella typhimurium* Strains and *E. coli* Strain

The histidine dependent strains are derived from *S. typhimurium* strain LT2 through a mutation in the histidine locus. Additionally due to the "deep rough" (*rfa*-minus) mutation they possess a faulty lipopolysaccharide envelope which enables substances to penetrate the cell wall more easily. A further mutation causes a reduction in the activity of an excision repair system. The latter alteration includes mutational processes in the nitrate reductase and biotin genes produced in a UV-sensitive area of the gene named "*uvrB*-minus". In the strains TA 98 and TA 100 the R-factor plasmid pKM 101 carries the ampicillin resistance marker (6).

Strain WP2 (4) and its derivatives all carry the same defect in one of the genes for tryptophan biosynthesis. Tryptophan-independent (*Trp*⁺) mutants (revertants) can arise either by a base change at the site of the original alteration or by a base change elsewhere in the chromosome so that the original defect is suppressed. This second possibility can occur in several different ways so that the system seems capable of detecting all types of mutagen which substitute one base for another. Additionally, the *uvrA* derivative is deficient in the DNA repair process (excision repair damage). Such a repair-deficient strain may be more readily mutated by agents.

When summarised the mutations of the TA strains and the *E. coli* strain, used in this study can be described as follows:

<i>Salmonella typhimurium</i>		
Strains	Genotype	Type of mutations indicated
TA 1537	his C 3076; <i>rfa</i> ⁻ ; <i>uvrB</i> ⁻	frame shift mutations
TA 98	his D 3052; <i>rfa</i> ⁻ ; <i>uvrB</i> ⁻ ; R-factor	" "
TA 1535	his G 46; <i>rfa</i> ⁻ ; <i>uvrB</i> ⁻	base-pair substitutions
TA 100	his G 46; <i>rfa</i> ⁻ ; <i>uvrB</i> ⁻ ; R-factor	" "
<i>Escherichia coli</i>		
WP2 <i>uvrA</i>	<i>trp</i> ⁻ ; <i>uvrA</i> ⁻	base-pair substitutions and others

Regular checking of the properties of the strains regarding the membrane permeability and ampicillin resistance as well as spontaneous mutation rates is performed in RCC Cytotest Cell Research according to B. Ames et al. (1) and D. Maron and B. Ames (6). In this way it was ensured that the experimental conditions set down by Ames were fulfilled.

The bacterial strains TA 1535, TA 1537, TA 98, TA 100, and WP2 *uvrA* were obtained from Trinova Biochem GmbH (35394 Gießen, Germany).

8.3.2 Storage

The strain cultures were stored as stock cultures in ampoules with nutrient broth + 5 % DMSO (MERCK, D-64293 Darmstadt) in liquid nitrogen.

8.3.3 Precultures

From the thawed ampoules of the strains 0.5 mL suspension was transferred into 250 mL Erlenmeyer flasks containing 20 mL nutrient medium. A solution of 20 µL ampicillin (25 µg/mL) was added to the strains TA 98 and TA 100. This nutrient medium contains per litre:

8 g Merck Nutrient Broth (MERCK, D-64293 Darmstadt)
5 g NaCl (MERCK, D-64293 Darmstadt)

The bacterial cultures were incubated in a shaking water bath for 4 hours at 37° C.

8.3.4 Selective Agar

The plates with the selective agar were obtained from E. Merck, D-64293 Darmstadt (Catalogue No.:1.13496.00.1; Lot No.:62355).

8.3.5 Overlay Agar

The overlay agar contains per litre:

for Salmonella strains:

6.0 g MERCK Agar Agar*
6.0 g NaCl*
10.5 mg L-Histidine×HCl×H₂O*
12.2 mg Biotin*

for Escherichia coli:

6.0 g MERCK Agar Agar*
6.0 g NaCl*
2.5 mg Tryptophan*

* (MERCK, D-64293 Darmstadt)

Sterilisations were performed at 121° C in an autoclave.

8.4 Mammalian Microsomal Fraction S9 Mix

The bacteria used in this assay do not possess the enzyme systems which, in mammals, are known to convert promutagens into active DNA damaging metabolites. In order to overcome this major drawback an exogenous metabolic system is added in form of mammalian microsome enzyme activation mixture.

8.4.1 S9 (Preparation by R C C - C C R)

Phenobarbital/ β -Naphthoflavone induced rat liver S9 is used as the metabolic activation system. The S9 is prepared from 8 - 12 weeks old male Wistar Hanlbm rats, weight approx. 220 - 320 g induced by applications of 80 mg/kg b.w. Phenobarbital i.p. (Desitin; D-22335 Hamburg) and β -Naphthoflavone p.o. (Aldrich, D-89555 Steinheim) each on three consecutive days. The livers are prepared 24 hours after the last treatment. The S9 fractions are produced by dilution of the liver homogenate with a KCl solution (1+3) followed by centrifugation at 9000 g. Aliquots of the supernatant are frozen and stored in ampoules at -80° C. Small numbers of the ampoules can be kept at -20°C for up to one week. Each batch of S9 mix is routinely tested with 2-aminoanthracene as well as benzo(a)pyrene.

The protein concentration in the S9 preparation was 38.1 mg/mL (lot no. R 031106) in both experiments.

8.4.2 S9 Mix

Before the experiment an appropriate quantity of S9 supernatant was thawed and mixed with S9 co-factor solution. The amount of S9 supernatant was 10% v/v in the S9 mix. Cofactors are added to the S9 mix to reach the following concentrations in the S9 mix:

8 mM $MgCl_2$
33 mM KCl
5 mM Glucose-6-phosphate
5 mM NADP

in 100 mM sodium-ortho-phosphate-buffer, pH 7.4.

During the experiment the S9 mix was stored in an ice bath. The S9 mix preparation was performed according to Ames et al.(1).

8.5 Pre-Experiment for Toxicity

To evaluate the toxicity of the test item a pre-experiment was performed with strains TA 1535, TA 1537, TA 98, TA 100, and WP2 uvrA. Eight concentrations were tested for toxicity and mutation induction with three plates each. The experimental conditions in this pre-experiment were the same as described below for the experiment I (plate incorporation test).

Toxicity of the test item results in a reduction in the number of spontaneous revertants or a clearing of the bacterial background lawn.

The pre-experiment is reported as main experiment I, if the following criteria are met:

Evaluable plates (>0 colonies) at five concentrations or more in all strains used.

8.6 Dose Selection

In the pre-experiment the concentration range of the test item was 3 – 5000 µg/plate. The pre-experiment is reported as experiment I since no relevant toxic effects were observed and 5000 µg/plate were chosen as maximal concentration.

The concentration range included two logarithmic decades. The following concentrations were tested in experiment II:

33; 100; 333; 1000; 2500; and 5000 µg/plate

8.7 Experimental Performance

For each strain and dose level including the controls, three plates were used.

The following materials were mixed in a test tube and poured onto the selective agar plates:

- 100 µL Test solution at each dose level, solvent (negative control) or reference mutagen solution (positive control),
- 500 µL S9 mix (for test with metabolic activation) or S9 mix substitution buffer (for test without metabolic activation),
- 100 µL Bacteria suspension (cf. test system, pre-culture of the strains),
- 2000 µL Overlay agar

In the pre-incubation assay 100 µL test solution, 500 µL S9 mix / S9 mix substitution buffer and 100 µL bacterial suspension were mixed in a test tube and shaken at 37° C for 60 minutes. After pre-incubation 2.0 mL overlay agar (45° C) was added to each tube. The mixture was poured on selective agar plates.

After solidification the plates were incubated upside down for at least 48 hours at 37° C in the dark (2).

8.8 Data Recording

The colonies were counted using the Petri Viewer Mk2 (Perceptive Instruments Ltd, Suffolk CB 7BN, UK) with the software program Ames Study Manager. The counter was connected to an IBM AT compatible PC with printer to print out the individual values and the means from the plates for each concentration together with standard deviations and enhancement factors as compared to the spontaneous reversion rates (see tables of results).

8.9 Acceptability of the Assay

The Salmonella typhimurium and Escherichia coli reverse mutation assay is considered acceptable if it meets the following criteria:

- regular background growth in the negative and solvent control
- the spontaneous reversion rates in the negative and solvent control are in the range of our historical data
- the positive control substances should produce a significant increase in mutant colony frequencies

8.10 Evaluation of Results

A test item is considered as a mutagen if a biologically relevant increase in the number of revertants exceeding the threshold of twice (strains TA 98, TA 100, and WP2 uvrA) or thrice (strains TA 1535 and TA 1537) the colony count of the corresponding solvent control is observed (3).

A dose dependent increase is considered biologically relevant if the threshold is exceeded at more than one concentration (2).

An increase exceeding the threshold at only one concentration is judged as biologically relevant if reproduced in an independent second experiment.

A dose dependent increase in the number of revertant colonies below the threshold is regarded as an indication of a mutagenic potential if reproduced in an independent second experiment. However, whenever the colony counts remain within the historical range of negative and solvent controls such an increase is not considered biologically relevant.

8.11 Biometry

According to the OECD guideline 471, a statistical analysis of the data is not mandatory.

9 DISCUSSION OF RESULTS

The test item Glyphosate technical (NUP-05067) was assessed for its potential to induce gene mutations in the plate incorporation test (experiment I) and the pre-incubation test (experiment II) using *Salmonella typhimurium* strains TA 1535, TA 1537, TA 98, and TA 100, and the *Escherichia coli* strain WP2 uvrA.

The assay was performed in two independent experiments both with and without liver microsomal activation. Each concentration and the controls were tested in triplicate. The test item was tested at the following concentrations:

Pre-Experiment/Experiment I: 3; 10; 33; 100; 333; 1000; 2500; and 5000 µg/plate

Experiment II: 33; 100; 333; 1000; 2500; and 5000 µg/plate

The plates incubated with the test item showed normal background growth in experiment I and II with and without metabolic activation. Only in experiment I in strain WP2 uvrA reduced background growth was observed at 5000 µg/plate in the absence of metabolic activation.

No toxic effects, evident as a reduction in the number of revertants, occurred in the test groups with and without metabolic activation. Minor toxic effects occurred at 5000 µg/plate in strain WP2 uvrA in the absence of metabolic activation in experiment I and II.

No substantial increase in revertant colony numbers of any of the five tester strains was observed following treatment with Glyphosate technical (NUP-05067) at any dose level, neither in the presence nor absence of metabolic activation (S9 mix). There was also no tendency of higher mutation rates with increasing concentrations in the range below the generally acknowledged border of biological relevance.

Appropriate reference mutagens were used as positive controls. They showed a distinct increase of induced revertant colonies.

In conclusion, it can be stated that during the described mutagenicity test and under the experimental conditions reported, the test item did not induce gene mutations by base pair changes or frameshifts in the genome of the strains used.

10 REFERENCES

1. Ames, B.N., J. McCann, and E. Yamasaki (1977)
Methods for detecting carcinogens and mutagens with the Salmonella/mammalian microsome mutagenicity test
In: B.J. Kilbey et al. (Eds.) "Handbook of Mutagenicity Test Procedures" Elsevier, Amsterdam, 1-17
2. de Serres F.J. and M.D. Shelby (1979)
Recommendations on data production and analysis using the Salmonella/microsome mutagenicity assay
Mutation Res. 64, 159-165
3. Hollstein, M., J. McCann, F.A. Angelosanto and W.W. Nichols (1979)
Short-term tests for carcinogens and mutagens
Mutation Res. 65, 133-226
4. Green, M.H.L. and W.J. Muriel (1976)
Mutagen Testing Using TRP⁺ Reversion in Escherichia Coli
Mutation. Res. 38, 3- 32
5. Maron D.M., J. Katzenellenbogen and B.N. Ames, (1981)
Compatibility of organic solvents with the Salmonella/Microsome Test
Mutation Res. 88, 343-350
6. Maron D.M., Ames, B.N. (1983)
Revised methods for the Salmonella mutagenicity test
Mutation Res. 113, 173-215

11 DISTRIBUTION OF THE REPORT

Sponsor	2 × (copy)
Contacting Institute	1 x (electronic copy (pdf-file))
Study Director	1 × (original)

12 SUMMARY OF RESULTS

12.1 Summary of Results Pre-Experiment/Experiment I

Study Name: 1061403
 Experiment: 1061403 VV Plate
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 18/01/2007
 Date Counted: 19/01/2007

Metabolic Activation	Test Group	Dose Level (µg/plate)	Revertant Colony Counts (Mean ±SD)				
			TA 1535	TA 1537	TA 98	TA 100	WP2 uvrA
Without Activation	Deionised water		20 ± 6	10 ± 4	24 ± 5	127 ± 8	53 ± 3
	Untreated		19 ± 8	11 ± 4	33 ± 6	139 ± 16	53 ± 10
	Glyphosate technical (NUP-05067)	3 µg	19 ± 4	9 ± 3	28 ± 6	126 ± 7	44 ± 7
		10 µg	20 ± 6	10 ± 2	28 ± 4	136 ± 11	54 ± 6
		33 µg	21 ± 2	11 ± 3	26 ± 10	128 ± 16	51 ± 10
		100 µg	20 ± 7	12 ± 4	25 ± 2	131 ± 8	51 ± 6
		333 µg	12 ± 3	14 ± 2	25 ± 5	118 ± 10	54 ± 4
		1000 µg	20 ± 8	8 ± 2	28 ± 7	119 ± 15	52 ± 5
		2500 µg	18 ± 5	9 ± 2	24 ± 3	135 ± 16	43 ± 4
		5000 µg	17 ± 6	9 ± 1	21 ± 7	116 ± 3	22 ± 6 ^R
	NaN3	10 µg	1991 ± 41		436 ± 43	2196 ± 97	
	4-NOPD	10 µg		102 ± 9			
	4-NOPD	50 µg					
	MMS	3.0 µL					1304 ± 54
With Activation	Deionised water		21 ± 2	15 ± 2	34 ± 4	138 ± 3	73 ± 4
	Untreated		21 ± 5	20 ± 4	41 ± 7	135 ± 2	72 ± 10
	Glyphosate technical (NUP-05067)	3 µg	23 ± 4	15 ± 5	33 ± 3	127 ± 7	69 ± 14
		10 µg	22 ± 3	16 ± 0	41 ± 9	139 ± 4	77 ± 2
		33 µg	20 ± 2	14 ± 3	34 ± 8	148 ± 9	74 ± 10
		100 µg	16 ± 7	12 ± 4	32 ± 7	148 ± 1	71 ± 14
		333 µg	24 ± 4	16 ± 1	35 ± 7	121 ± 6	76 ± 11
		1000 µg	21 ± 9	19 ± 3	38 ± 2	144 ± 8	67 ± 4
		2500 µg	19 ± 2	15 ± 4	34 ± 2	137 ± 9	60 ± 9
		5000 µg	22 ± 3	18 ± 3	28 ± 6	112 ± 8	55 ± 7
	2-AA	2.5 µg	345 ± 11	208 ± 9	1459 ± 133	2023 ± 40	
	2-AA	10.0 µg					322 ± 16

Key to Positive Controls

NaN3	sodium azide
2-AA	2-aminoanthracene
4-NOPD	4-nitro-o-phenylene-diamine
MMS	methyl methane sulfonate

Key to Plate Postfix Codes

R	Reduced background growth
---	---------------------------

12.2 Summary of Results Experiment II

Study Name: 1061403
 Experiment: 1061403 HV2 Pre
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 22/01/2007
 Date Counted: 25/01/2007

Metabolic Activation	Test Group	Dose Level (µg/plate)	Revertant Colony Counts (Mean ±SD)				
			TA 1535	TA 1537	TA 98	TA 100	WP2 uvrA
Without Activation	Deionised water		16 ± 7	8 ± 2	32 ± 3	150 ± 2	58 ± 5
	Untreated		12 ± 2	11 ± 4	28 ± 7	124 ± 27	52 ± 3
	Glyphosate technical	33 µg	19 ± 4	13 ± 4	28 ± 6	131 ± 8	54 ± 2
	(NUP-05067)	100 µg	16 ± 6	14 ± 1	26 ± 3	144 ± 7	54 ± 11
		333 µg	15 ± 5	11 ± 4	30 ± 4	147 ± 3	49 ± 12
		1000 µg	13 ± 4	10 ± 4	30 ± 4	148 ± 8	41 ± 7
		2500 µg	19 ± 3	10 ± 1	24 ± 3	135 ± 3	30 ± 4
		5000 µg	10 ± 3	9 ± 2	25 ± 9	105 ± 13	24 ± 11
		NaN3	10 µg	2045 ± 99		2246 ± 39	
		4-NOPD	10 µg			356 ± 28	
	4-NOPD	50 µg			105 ± 5		
	MMS	3.0 µL			705 ± 40		
With Activation	Deionised water		28 ± 2	18 ± 3	41 ± 4	164 ± 2	66 ± 10
	Untreated		21 ± 2	24 ± 2	40 ± 9	153 ± 16	68 ± 17
	Glyphosate technical	33 µg	24 ± 4	21 ± 4	44 ± 10	167 ± 14	68 ± 18
	(NUP-05067)	100 µg	22 ± 8	19 ± 1	36 ± 14	161 ± 11	62 ± 4
		333 µg	23 ± 9	18 ± 4	33 ± 3	166 ± 19	73 ± 9
		1000 µg	21 ± 2	19 ± 4	39 ± 9	154 ± 32	67 ± 7
		2500 µg	19 ± 8	14 ± 4	39 ± 13	153 ± 11	49 ± 9
		5000 µg	21 ± 2	19 ± 5	42 ± 8	146 ± 34	49 ± 5
		2-AA	2.5 µg	335 ± 12	229 ± 45	1008 ± 128	2331 ± 280
		2-AA	10.0 µg			287 ± 18	

Key to Positive Controls

NaN3	sodium azide
2-AA	2-aminoanthracene
4-NOPD	4-nitro-o-phenylene-diamine
MMS	methyl methane sulfonate

13 HISTORICAL CONTROL DATA

These data represent the laboratory's historical control data from May 2005 until June 2006 representing approx. 200 experiments (WP2 uvrA the historical data are based on approx. 100 experiments).

Strain		without S9 mix				with S9 mix			
		Mean	SD	Min	Max	Mean	SD	Min	Max
TA 1535	Solvent control	20.8	4.7	9	35	24.7	5.9	7	43
	Negative control	20.4	4.4	11	31	24.2	5.5	10	38
	Positive control	1422.0	464.7	781	4900	332.0	95.3	107	695
TA1537	Solvent control	11.2	3.7	5	28	16.2	5.0	6	36
	Negative control	11.6	4.0	4	28	17.1	5.4	7	34
	Positive control	99.8	32.5	53	425	276.8	132.6	59	746
TA 98	Solvent control	28.1	6.1	15	49	37.9	7.4	20	57
	Negative control	30.2	6.6	16	60	39.0	7.5	18	64
	Positive control	439.0	155.2	176	1818	1839.4	898.6	407	4891
TA 100	Solvent control	130.7	20.8	87	197	147.0	25.5	84	255
	Negative control	138.2	21.6	86	216	150.1	24.2	96	214
	Positive control	2083.1	281.3	616	2872	2372.9	958.4	417	5230
WP2uvrA	Solvent control	55.8	7.2	31	74	63.9	9.1	34	84
	Negative control	55.9	8.6	36	76	65.6	10.4	33	91
	Positive control	991.0	522.9	249	1810	319.4	84.8	211	930

Mean = mean value of revertants/plate

SD = standard deviation

Min = minimal value/Max = maximal value

14 ANNEX I: TABLES OF RESULTS (8 PAGES)

Pre-Experiment and Experiment I: 1061403 VV Plate Incorporation (4 pages)

Experiment II: 1061403 Pre-Incubation (4 pages)

This document is not the property of EFSA and is provided for giving full effect to the right of public access to documents under EU law.
The document may be subject to rights such as intellectual property and copy rights of third parties.
Furthermore, this document may fall under a regulatory data protection regime.
Consequently, any publication, distribution, reproduction and/or publishing and any commercial exploitation and use
of this document or its contents without the permission of the owner
of this document may therefore be prohibited and violate the rights of its owner.

Study Name: 1061403
 Experiment: 1061403 VV Plate
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 16/01/2007
 Date Counted: 19/01/2007

Without metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Glyphosate technical (NUP- 05067)	3 µg	19.3	3.5	1.0	23, 16, 19
		10 µg	19.7	5.5	1.0	17, 16, 26
		33 µg	20.7	1.5	1.1	21, 19, 22
		100 µg	19.7	6.7	1.0	12, 23, 24
		333 µg	12.0	3.5	0.6	10, 10, 16
		1000 µg	19.7	8.1	1.0	14, 29, 16
		2500 µg	18.3	5.1	0.9	17, 14, 24
		5000 µg	17.0	6.1	0.9	21, 20, 10
	Deionised water		19.7	5.5		26, 16, 17
	Untreated Control		19.0	8.2		17, 28, 12
TA 1537	Glyphosate technical (NUP- 05067)	3 µg	9.3	3.1	0.9	10, 6, 12
		10 µg	10.0	2.0	1.0	10, 8, 12
		33 µg	11.3	3.2	1.1	10, 9, 15
		100 µg	12.3	4.0	1.2	10, 17, 10
		333 µg	13.7	1.5	1.4	12, 14, 15
		1000 µg	8.3	2.1	0.8	10, 6, 9
		2500 µg	8.7	1.5	0.9	10, 9, 7
		5000 µg	9.0	1.0	0.9	8, 9, 10
	Deionised water		10.0	3.6		14, 7, 9
	Untreated Control		11.3	4.2		16, 8, 10
TA 98	Glyphosate technical (NUP- 05067)	3 µg	28.0	6.2	1.2	35, 26, 23
		10 µg	27.7	4.2	1.2	29, 23, 31
		33 µg	25.7	10.4	1.1	29, 14, 34
		100 µg	25.0	1.7	1.1	26, 23, 26
		333 µg	25.3	4.9	1.1	22, 23, 31
		1000 µg	27.7	7.0	1.2	27, 21, 35
		2500 µg	24.3	3.2	1.0	28, 22, 23
		5000 µg	20.7	6.7	0.9	15, 28, 19
	Deionised water		23.7	5.0		23, 19, 29
	Untreated Control		32.7	6.4		28, 40, 30
TA 100	Glyphosate technical (NUP- 05067)	3 µg	126.3	6.7	1.0	119, 128, 132
		10 µg	136.3	11.0	1.1	149, 130, 130
		33 µg	128.3	15.6	1.0	145, 114, 126
		100 µg	131.0	7.9	1.0	122, 134, 137
		333 µg	117.7	10.3	0.9	115, 129, 109
		1000 µg	119.0	15.4	0.9	136, 115, 108
		2500 µg	134.7	16.2	1.1	144, 144, 116
		5000 µg	116.0	2.6	0.9	119, 114, 115
	Deionised water		127.3	8.0		119, 135, 128
	Untreated Control		139.0	15.6		121, 147, 149

Study Name: 1061403
 Experiment: 1061403 VV Plate
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 16/01/2007
 Date Counted: 19/01/2007

Without metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
WP2 uvrA	Glyphosate technical (NUP-05067)	3 µg	44.0	6.9	0.8	40, 40, 52
		10 µg	53.7	5.5	1.0	59, 54, 48
		33 µg	50.7	9.8	1.0	62, 45, 45
		100 µg	50.7	5.5	1.0	48, 47, 57
		333 µg	54.0	4.0	1.0	54, 58, 50
		1000 µg	52.3	4.5	1.0	57, 48, 52
		2500 µg	43.3	4.2	0.8	40, 48, 42
		5000 µg	22.0	5.6	0.4	17 R, 21 R, 28 R
	Deionised water		53.0	3.5		55, 55, 49
	Untreated Control		53.3	9.7		64, 51, 45
TA 1535	NaN3	10 µg	1991.0	40.5	101.2	1992, 2031, 1950
TA 1537	4-NOPD	50 µg	102.0	9.0	10.2	93, 111, 102
TA 98	4-NOPD	10 µg	436.0	43.2	18.4	445, 474, 389
TA 100	NaN3	10 µg	2195.7	97.0	17.2	2259, 2084, 2244
WP2 uvrA	MMS	3.0 µL	1303.7	54.1	24.6	1343, 1326, 1242

Key to Positive Controls

Key to Plate Postfix Codes

NaN3 sodium azide
 4-NOPD 4-nitro-o-phenylene-diamine
 MMS methyl methane sulfonate

R Reduced background growth

Study Name: 1061403
 Experiment: 1061403 VV Plate
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 16/01/2007
 Date Counted: 19/01/2007

With metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Glyphosate technical (NUP- 05067)	3 µg	22.7	4.0	1.1	22, 19, 27
		10 µg	22.3	3.2	1.0	21, 26, 20
		33 µg	20.3	1.5	1.0	20, 19, 22
		100 µg	16.3	6.7	0.8	24, 13, 12
		333 µg	24.3	4.2	1.1	29, 21, 23
		1000 µg	21.3	9.0	1.0	12, 30, 22
		2500 µg	19.3	2.1	0.9	21, 17, 20
		5000 µg	21.7	2.5	1.0	24, 22, 19
	Deionised water Untreated Control		21.3	1.5		21, 20, 23
	Glyphosate technical (NUP- 05067)	3 µg	14.7	4.5	1.0	15, 19, 10
		10 µg	16.0	0.0	1.0	16, 16, 16
		33 µg	13.7	3.2	0.9	10, 15, 16
		100 µg	12.3	4.0	0.8	10, 10, 17
		333 µg	16.0	1.0	1.0	17, 15, 16
		1000 µg	19.0	3.5	1.2	15, 21, 21
		2500 µg	15.3	3.5	1.0	12, 19, 15
		5000 µg	18.3	3.2	1.2	22, 17, 16
	Deionised water Untreated Control		15.3	1.5		15, 17, 14
	Glyphosate technical (NUP- 05067)	3 µg	32.7	2.5	1.0	35, 30, 33
		10 µg	40.7	8.7	1.2	31, 48, 43
		33 µg	34.0	7.5	1.0	41, 26, 35
		100 µg	31.7	6.7	0.9	35, 24, 36
		333 µg	35.0	7.0	1.0	40, 38, 27
		1000 µg	38.0	2.0	1.1	40, 38, 36
		2500 µg	34.3	1.5	1.0	33, 34, 36
		5000 µg	28.0	6.2	0.8	35, 23, 26
	Deionised water Untreated Control		33.7	4.2		29, 37, 35
	Glyphosate technical (NUP- 05067)	3 µg	127.0	6.6	0.9	126, 134, 121
		10 µg	139.0	4.4	1.0	134, 141, 142
		33 µg	148.0	8.7	1.1	142, 144, 158
		100 µg	148.0	1.0	1.1	149, 148, 147
		333 µg	120.7	6.4	0.9	118, 128, 116
		1000 µg	144.3	8.3	1.0	147, 135, 151
		2500 µg	137.0	9.2	1.0	129, 147, 135
		5000 µg	111.7	8.1	0.8	121, 107, 107
	Deionised water Untreated Control		138.3	2.9		140, 140, 135
	Glyphosate technical (NUP- 05067)	3 µg	127.0	6.6	0.9	126, 134, 121
		10 µg	139.0	4.4	1.0	134, 141, 142
		33 µg	148.0	8.7	1.1	142, 144, 158
		100 µg	148.0	1.0	1.1	149, 148, 147
		333 µg	120.7	6.4	0.9	118, 128, 116
		1000 µg	144.3	8.3	1.0	147, 135, 151
		2500 µg	137.0	9.2	1.0	129, 147, 135
		5000 µg	111.7	8.1	0.8	121, 107, 107
	Deionised water Untreated Control		138.3	2.9		140, 140, 135
	Glyphosate technical (NUP- 05067)	3 µg	127.0	6.6	0.9	126, 134, 121
		10 µg	139.0	4.4	1.0	134, 141, 142
		33 µg	148.0	8.7	1.1	142, 144, 158
		100 µg	148.0	1.0	1.1	149, 148, 147
		333 µg	120.7	6.4	0.9	118, 128, 116
		1000 µg	144.3	8.3	1.0	147, 135, 151
		2500 µg	137.0	9.2	1.0	129, 147, 135
		5000 µg	111.7	8.1	0.8	121, 107, 107
	Deionised water Untreated Control		138.3	2.9		140, 140, 135

Study Name: 1061403
 Experiment: 1061403 VV Plate
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 16/01/2007
 Date Counted: 19/01/2007

With metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
WP2 uvrA	Glyphosate technical (NUP- 05067)	3 µg	69.3	13.8	0.9	85, 64, 59
		10 µg	77.3	1.5	1.1	79, 76, 77
		33 µg	74.3	9.7	1.0	72, 66, 85
		100 µg	70.7	14.3	1.0	74, 83, 55
		333 µg	76.3	11.2	1.0	64, 79, 86
		1000 µg	67.3	4.2	0.9	66, 72, 64
		2500 µg	60.3	9.1	0.8	64, 50, 67
		5000 µg	55.0	6.9	0.8	59, 59, 47
	Deionised water		73.0	3.6		74, 76, 69
	Untreated Control		71.7	9.7		74, 80, 61
TA 1535	2-AA	2.5 µg	345.0	10.5	16.2	355, 334, 346
TA 1537	2-AA	2.5 µg	208.0	9.2	13.6	206, 200, 218
TA 98	2-AA	2.5 µg	1450.3	132.5	43.3	1523, 1548, 1307
TA 100	2-AA	2.5 µg	2023.3	39.7	14.6	2069, 2004, 1997
WP2 uvrA	2-AA	10.0 µg	322.3	16.3	4.4	335, 304, 328

Key to Positive Controls

2-AA 2-aminoanthracene

Study Name: 1061403
 Experiment: 1061403 HV2 Pre
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 22/01/2007
 Date Counted: 25/01/2007

Without metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Glyphosate technical (NUP- 05067)	33 µg	18.7	3.5	1.2	22, 15, 19
		100 µg	15.7	6.0	1.0	15, 22, 10
		333 µg	15.3	5.0	1.0	16, 20, 10
		1000 µg	12.7	4.0	0.8	8, 15, 15
		2500 µg	19.0	2.6	1.2	21, 20, 16
		5000 µg	10.3	2.5	0.7	10, 13, 8
	Deionised water Untreated Control		15.7 12.3	7.1 2.1		22, 17, 8 13, 14, 10
TA 1537	Glyphosate technical (NUP- 05067)	33 µg	13.3	3.5	1.7	10, 17, 13
		100 µg	14.0	1.0	1.8	14, 15, 13
		333 µg	10.7	3.8	1.4	8, 15, 9
		1000 µg	9.7	4.0	1.3	6, 14, 9
		2500 µg	9.7	0.6	1.3	10, 9, 10
		5000 µg	8.7	1.5	1.1	10, 7, 9
	Deionised water Untreated Control		7.7 11.3	2.3 4.2		9, 9, 5 16, 10, 8
TA 98	Glyphosate technical (NUP- 05067)	33 µg	28.0	6.0	0.9	28, 22, 34
		100 µg	26.3	2.5	0.8	29, 26, 24
		333 µg	30.0	4.4	0.9	28, 35, 27
		1000 µg	30.3	4.2	0.9	27, 35, 29
		2500 µg	24.3	2.5	0.8	24, 27, 22
		5000 µg	24.7	8.6	0.8	23, 34, 17
	Deionised water Untreated Control		32.3 28.3	3.1 6.5		33, 35, 29 28, 35, 22
TA 100	Glyphosate technical (NUP- 05067)	33 µg	131.0	7.8	0.9	122, 136, 135
		100 µg	144.3	7.0	1.0	151, 137, 145
		333 µg	146.7	2.9	1.0	150, 145, 145
		1000 µg	147.7	8.1	1.0	157, 144, 142
		2500 µg	134.7	2.5	0.9	132, 135, 137
		5000 µg	105.0	13.5	0.7	116, 90, 109
	Deionised water Untreated Control		150.3 124.3	1.5 27.4		150, 152, 149 108, 109, 156

Study Name: 1061403
Experiment: 1061403 HV2 Pre
Assay Conditions:

Study Code: RCC-CCR 1061403
Date Plated: 22/01/2007
Date Counted: 25/01/2007

Without metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
WP2 uvrA	Glyphosate technical (NUP-05067)	33 µg	54.3	2.1	0.9	52, 55, 56
		100 µg	54.3	11.0	0.9	65, 43, 55
		333 µg	49.3	11.8	0.9	42, 63, 43
		1000 µg	40.7	6.7	0.7	33, 45, 44
		2500 µg	30.0	3.6	0.5	29, 27, 34
		5000 µg	24.3	11.2	0.4	20, 37, 16
	Deionised water		57.7	4.7		56, 54, 63
	Untreated Control		52.0	3.0		55, 52, 49
TA 1535	NaN3	10 µg	2045.3	99.4	130.6	2125, 2077, 1934
TA 1537	4-NOPD	50 µg	104.7	4.5	13.7	100, 109, 105
TA 98	4-NOPD	10 µg	356.3	28.2	11.0	376, 369, 324
TA 100	NaN3	10 µg	2246.3	39.1	14.9	2243, 2209, 2287
WP2 uvrA	MMS	3.0 µL	705.3	40.0	12.2	746, 666, 704

Key to Positive Controls

NaN3 sodium azide
4-NOPD 4-nitro-o-phenylene-diamine
MMS methyl methane sulfonate

Study Name: 1061403
 Experiment: 1061403 HV2 Pre
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 22/01/2007
 Date Counted: 25/01/2007

With metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Glyphosate technical (NUP-05067)	33 µg	23.7	4.0	0.8	23, 20, 28
		100 µg	21.7	7.8	0.8	13, 24, 28
		333 µg	22.7	9.3	0.8	12, 27, 29
		1000 µg	21.0	1.7	0.7	20, 23, 20
		2500 µg	18.7	7.5	0.7	23, 23, 10
		5000 µg	20.7	1.5	0.7	22, 21, 19
	Deionised water		28.3	2.1		26, 29, 30
	Untreated Control		21.3	2.1		19, 22, 23
TA 1537	Glyphosate technical (NUP-05067)	33 µg	20.7	4.2	1.1	24, 16, 22
		100 µg	19.3	0.6	1.1	19, 19, 20
		333 µg	17.7	4.0	1.0	13, 20, 20
		1000 µg	19.0	4.4	1.0	22, 21, 14
		2500 µg	14.3	3.8	0.8	10, 16, 17
		5000 µg	18.7	4.5	1.0	23, 14, 19
	Deionised water		18.3	2.9		20, 20, 15
	Untreated Control		23.7	2.1		26, 23, 22
TA 98	Glyphosate technical (NUP-05067)	33 µg	44.0	10.4	1.1	38, 38, 56
		100 µg	36.3	13.6	0.9	47, 41, 21
		333 µg	32.7	3.2	0.8	29, 35, 34
		1000 µg	39.0	8.5	0.9	38, 48, 31
		2500 µg	39.0	13.1	0.9	54, 33, 30
		5000 µg	42.3	8.0	1.0	43, 50, 34
	Deionised water		41.3	3.5		38, 41, 45
	Untreated Control		40.0	8.7		34, 50, 36
TA 100	Glyphosate technical (NUP-05067)	33 µg	166.7	13.7	1.0	151, 176, 173
		100 µg	161.3	10.8	1.0	149, 166, 169
		333 µg	166.3	18.5	1.0	176, 145, 178
		1000 µg	154.0	32.4	0.9	125, 148, 189
		2500 µg	152.7	11.1	0.9	141, 154, 163
		5000 µg	146.0	33.8	0.9	182, 141, 115
	Deionised water		163.7	2.3		165, 165, 161
	Untreated Control		152.7	15.5		159, 135, 164

Study Name: 1061403
 Experiment: 1061403 HV2 Pre
 Assay Conditions:

Study Code: RCC-CCR 1061403
 Date Plated: 22/01/2007
 Date Counted: 25/01/2007

With metabolic activation

Strain	Compound	Dose level per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
WP2 uvrA	Glyphosate technical (NUP- 05067)	33 µg	68.0	18.2	1.0	47, 79, 78
		100 µg	62.0	3.6	0.9	59, 61, 66
		333 µg	73.0	8.7	1.1	67, 83, 69
		1000 µg	67.3	6.5	1.0	67, 61, 74
		2500 µg	49.3	8.6	0.8	40, 51, 57
		5000 µg	49.0	5.3	0.7	47, 45, 55
	Deionised water Untreated Control		65.7 68.0	10.4 16.6		54, 74, 69 56, 61, 87
TA 1535	2-AA	2.5 µg	335.0	12.1	11.8	337, 346, 322
TA 1537	2-AA	2.5 µg	228.7	44.5	12.5	205, 201, 280
TA 98	2-AA	2.5 µg	1008.3	128.5	24.4	1067, 861, 1097
TA 100	2-AA	2.5 µg	2330.7	279.7	14.2	2021, 2406, 2565
WP2 uvrA	2-AA	10.0 µg	286.7	17.6	4.4	292, 267, 301
Key to Positive Controls						
2-AA	2-aminoanthracene					

15 ANNEX II: COPY OF GLP CERTIFICATE OF ANALYSIS



VRéf :

CERTIFICAT D'ANALYSE
CERTIFICATE OF ANALYSIS

NRéf :

Nom du produit
Product name : Glyphosate Technical (NUP 05067)

Réf. lot
Batch ref. : 0608-1

Réf. d'analyse
Analytical ref. : N° 00200

Date d'analyse
Analytical date : November 06, 2006

Date de fabrication
Date of Manufacture : August 15, 2006

Date de péremption
Expiry date : August 15, 2008

Caractéristiques <i>Characteristics</i>	Réf. Méthode <i>Test method ref.</i>	Unités <i>Units</i>	Résultats <i>Results</i>
Aspect <i>Appearance</i>	Visual <i>Visual</i>	-	Poudre cristalline blanche <i>White crystal powder</i>
Glyphosate	CIPAC MT 284/TC(87)	%	95.0

Ce produit a été analysé en conformité avec les principes des Bonnes Pratiques de Laboratoire. Les données brutes relatives à l'analyse de cet échantillon sont archivées chez Nufarm SAS, Laboratoire de Chimie Analytique, Gagneville, France.
This product was analysed in compliance with Good Laboratory Practice standards. The deviation and characterization data for this material are located at Nufarm SAS, Analytical Chemistry Laboratory, Gagneville, France.

Date : 06 novembre 2006
Date : November 06, 2006

Date de la copie certifiée :
Date of certified copy : November 15, 2006

Approbateur du certificat :
Certificate approver:

Opérateur :
Operator:

Version n°3 du 01/02/02

DQERL9.110



Nufarm S.A.S.
28, boulevard Camélinat - BP 75 - 92233 GENNEVILLIERS Cedex (France)
Tél : 01 40 85 50 50 - Télécopieur : 01 47 92 25 45

Société par Actions Simplifiée au capital de 5 664 700 Euros
R.C.S. Nanterre 553 029 020 - SIRET 553 029 020 00028 - N° TVA CEE : FR-08552029028

GALLON 27500

Notre-Dame-de-la-Garenne
Tél : 02 32 64 74 00
Télécopieur : 02 32 33 03 52

16 ANNEX III: COPY OF GLP CERTIFICATE

Hessisches Ministerium für Umwelt,
ländlichen Raum und Verbraucherschutz

HESSEN



Gute Laborpraxis/Good Laboratory Practice

GLP-Bescheinigung/Statement of GLP Compliance
(gemäß/according to § 19b Abs. 1 Chemikaliengesetz)

Eine GLP-Inspektion zur Überwachung der Einhaltung
der GLP-Grundsätze gemäß Chemikaliengesetz bzw.
Richtlinie 88/320/EG wurde durchgeführt in

Assessment of conformity with GLP according to
Chemikaliengesetz and Directive 88/320/EEC at

☒ Prüfeinrichtung/Test facility

☐ Prüfstandort/Test site

RCC Cytotest Cell Research GmbH
RCC Cytotest Cell Research GmbH
In den Leppsteinwiesen 19
64380 Roldorf

(Unverwechselbare Bezeichnung und Adresse/Unequivocal name and address)

Prüfungen nach Kategorien/Areas of Expertise
(gemäß/according chemVwV-GLP Nr. 5.3/OECD guidance)

2 Prüfungen zur Bestimmung der toxiologischen
Eigenschaften
3 Prüfungen zur Bestimmung der erbgutverändernden
Eigenschaften (in vitro und in vivo)
8 Analytische Prüfungen an biologischen Materialien
9 Virussicherheitsprüfungen

2 Toxicity studies
3 Mutagenicity studies
8 Analytical studies on biological materials
9 Virus validation studies

03.06.; 19.07.-22.07.2004

Datum der Inspektion/Date of Inspection
(Tag Monat Jahr/day month year)

Die genannte Prüfeinrichtung befindet sich im nation-
alen GLP-Überwachungsverfahren und wird regel-
mäßig auf Einhaltung der GLP-Grundsätze überwacht.

The above mentioned test facility is included
in the national GLP Compliance Programme and is
inspected on a regular basis.

Auf der Grundlage des Inspektionsberichtes wird hiernit
bestätigt, dass in dieser Prüfeinrichtung die oben ge-
nannten Prüfungen unter Einhaltung der GLP- Grund-
sätze durchgeführt werden können.

Based on the inspection report it can be confirmed,
that this test facility is able to conduct the
aforementioned studies in compliance with the
Principles of GLP.

Im Auftrag

Referent, Wiesbaden, den Wiesbaden, den 06. Januar 2005
(Name und Funktion der verantwortlichen Person/
Name and function of responsible person)

Hess. Ministerium für Umwelt, ländlichen Raum und Verbraucherschutz,
Mainzer Straße 89 D65189 Wiesbaden

(Name und Adresse der GLP-Überwachungsbehörde/Name and address of the GLP Monitoring Authority)

D-65189 Wiesbaden, Mainzer Straße 89
Telefon: 0611. 81 50
Telefax: 0611. 81 51 54 1
E-Mail: poststelle@hmuiv.hessen.de

D-65187 Wiesbaden, Hölderlinstraße 1-3
Telefon: 0611. 81 70
Telefax: 0611. 81 72 18 1

